

Effects of pH, temperature and oxygen-limited condition on the virulence of *Vibrio parahaemolyticus*

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ABSTRACT

Introduction: *Vibrio parahaemolyticus* is a popular Gram-negative bacterium in the marine and estuarine regions. It can cause Early Mortality Syndrome (EMS), now named Acute Hepatopancreatic Necrosis Disease (AHPND), which resulted in severe losses to the shrimp culture. This study aimed to investigate the effect of pH, temperature, and oxygen-limited condition on the extracellular enzymatic activity of *V. parahaemolyticus*. **Methods:** *V. parahaemolyticus* XN9, an AHPND-causing strain, was cultured in Brain Heart Infusion (BHI) medium at different pHs (7.5, 8.0, 8.5 and 9.0), temperatures (25°C, 30°C, and 35°C) and different oxygen conditions (either 120rpm shaking or static with the presence of oxygen absorber packages). The activity of five extracellular enzymes, including caseinase, lecithinase, chitinase, gelatinase, and lipase, was assessed using the agar-based method with the corresponding media. **Results:** When pH was increased from 7.5 to 9.0, caseinase and lipase activity was decreased significantly by 88% and 44%. In contrast, gelatinase activity increased markedly from 0 to 1.38 ± 0.17 (+) mm, and lecithinase reached the highest activity, which was 2.96 ± 0.13 mm (++) at pH 8.5. Regarding effect of temperature, highest activity of caseinase (0.85 ± 0.13 mm (+)) and gelatinase (1.37 ± 0.25 mm (+)) was obtained at 35°C, lecithinase at 30°C and lipase at 25°C. Regarding the effect of oxygen level, the activity of most tested enzymes decreased significantly following the decrease of oxygen level. The highest activity of caseinase, gelatinase, and lipase was observed when the bacteria were cultured and tested in a fully oxygenated condition while lecithinase showed the highest activity when the bacteria were cultured in oxygenated condition but tested in oxygen-limited condition. No chitinase activity was observed in any of the tested conditions. **Conclusion:** Our data suggested that extracellular enzymatic activity of *V. parahaemolyticus* is significantly influenced by environmental conditions. No particular testing condition resulted in the highest activity for all tested enzymes. However, warm temperature (30/ 35°C), mildly alkaline pH (pH 8.0), and fully oxygenated condition could increase the overall extracellular enzymatic activity of *V. parahaemolyticus*, thus increase its potential virulence.

Key words: *Vibrio parahaemolyticus*, Acute Hepatopancreatic Necrosis Disease (AHPND), Early Mortality Syndrome (EMS), extracellular enzymatic activity, oxygen-limited, pH, temperature, virulence

INTRODUCTION

Vibrio parahaemolyticus is a halophilic Gram-negative bacterium. It lives ubiquitously as a free-living organism in the marine environment or a colonizer of many different kinds of marine organisms¹. This motile, curved shaped bacterium, is a well-known causative agent of food-borne acute gastroenteritis in humans due to the consumption of raw or undercooked seafood²⁻⁴. *V. parahaemolyticus* is also known to cause Early Mortality Syndrome (EMS) or Acute Hepatopancreatic Necrosis Disease (AHPND), which affects penaeid shrimp, causing massive death in larvae and young adults^{5,6}. In recent years, AHPND has brought devastating effects to the shrimp industry of various countries such as China, Vietnam, Malaysia, Philippines, Thailand, and

Mexico⁷⁻⁹. AHPND pathogenesis is mainly caused by a binary toxin PirA/B encoded on a plasmid in *V. parahaemolyticus*¹⁰. However, whether other toxins may also take part in causing this disease is still under investigation. The potential of a pathogen to cause so-called disease virulence reflects its ability to colonize, invade, escape the immune system, and obtain nutrition from the host. An important part of bacterial virulence is the ability to produce and secrete extracellular enzymes to break down and digest nutrients from the environment^{11,12}. However, the production of these enzymes is highly influenced by environmental factors such as nutrient supplement, dissolved oxygen, pH, temperature...¹³. In this study, the activity of five extracellular enzymes, including caseinase, lecithinase, gelatinase, lipase, and chitinase, was examined under the different pHs,

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temperatures and oxygen levels.

MATERIALS- METHODS

Bacteria strain

Vibrio parahaemolyticus XN9, an AHPND-causing isolate, was kindly provided by Nha Trang University¹⁴. It was streaked from glycerol stock on Thiosulfate-citrate-bile salts-sucrose agar (TCBS, Himedia, India). One colony was picked up for overnight culture in BHI medium at optimal culture condition (pH 8.5, 2.5% NaCl, 30°C, and static condition) described previously (14). For assessing the effect of culture condition on the activity of extracellular enzymes, pH, temperature, and oxygen level were adjusted around the optimal culture condition (pH 8.5, 2.5% NaCl, 30°C, and static condition). So, the testing conditions were 7.5, 8.0, 8.5 and 9.0 for pH; 25°C, 30°C and 35°C for temperature, and 120 rpm shaking or static condition with the presence of Oxygen absorber package (O-Buster, Hsiao Sung, Non-Oxygen Chemical Co. Ltd., China) adhered to the bottom side of the falcon cap for oxygen level.

Extracellular enzymatic testing

Egg-yolk agar containing 1mL of Egg Yolk Emulsion (Himedia), Tributyrin Agar (Himedia) with added Tributyrin (Himedia) (10mL/L), BHI agar plates containing 1.5% (w/v) skim milk, 8% gelatin and 2% (w/v) colloidal chitin was used for lecithinase, lipase, caseinase, gelatinase, and chitinase, respectively. The colloidal chitin was prepared, as previously described¹⁵. The overnight culture of *V. parahaemolyticus* was adjusted to OD_{600nm} of 0.08- 0.1, then 10 μ L of this bacterial suspension was dropped onto agar plates corresponding to the tested enzymes mentioned above. In the case of assessing the effect of oxygen, the overnight culture was dropped on the plate either without or with AnaeroPack® (Mitsubishi Gas Chemical, Japan) and plastic wrap. Positive controls used in these tests included *Staphylococcus aureus* ATCC29213 for caseinase and lipase, *Vibrio cholerae* for lecithinase and gelatinase and *Vibrio alginolyticus* for chitinase. After inoculation, the plates were incubated 24 hours for caseinase, gelatinase, and lipase and 48 hours for lecithinase and chitinase. For gelatinase, before reading the result, the agar plate was flooded with saturated ammonium sulfate ((NH₄)₂SO₄) to precipitate the undegraded gelatin. Clear halos surrounding the bacterial drop indicated the activity of the tested enzymes¹⁶⁻¹⁸. All the tests were triplicated.

Data analysis

Enzyme activity (EA) was calculated using the formula: where D is the diameter of the bacterial drop plus the clear halo zone (mm), and d is the diameter of the bacterial drop itself (mm). It is graded (-) if there was no visible hydrolytic area; (+) if the EA value is less than 2 mm and (++) if equal or higher than 2 mm (15). Each test was triplicated, and the obtained data were analyzed using two-way ANOVA (Excel software, Microsoft 7)¹⁴.

RESULTS

Effects of pH on extracellular enzymatic activities of *V. parahaemolyticus*

Following the increase of pH from 7.5 to 9.0, caseinase activity decreased significantly by nearly 88% from 3.55 ± 0.25 (++) to 0.41 ± 0.08 mm (+) and lipase activity decreased by roughly 44%, from 1.83 ± 0.29 (+) to 1.21 ± 0.25 mm (+). In contrast, there was a significant increase of gelatinase activity from an undetectable level at pH 7.5 to 1.38 ± 0.17 mm (+) at pH 9.0. On the other hand, lecithinase activity of *V. parahaemolyticus* was recorded as strong (++) in all tested pHs with the highest value obtained at pH 8.5 (2.96 ± 0.13 mm). Chitinase activity was not observed in any tested pHs (Figure 1, Table 1). In the increasing pH from 7.5 to 9.0, *V. parahaemolyticus* exhibited significant differences in enzymatic activities between four pH levels (p-value < 0.05, Supplementary Table 2 A).

Effects of temperature on extracellular enzymatic activities of *V. parahaemolyticus*

The rise of temperature from 25°C to 35°C led to strong decomposition of gelatin in the BHI medium with EA value increased by 56% from 0.86 ± 0.14 (+) to 1.37 ± 0.25 mm (+). It also resulted in slight increase of caseinase from 0.67 ± 0.10 (+) to 0.85 ± 0.13 mm (+). At 30°C, the obtained EA value was highest for lecithinase (2.96 ± 0.13 mm (++) but lowest for lipase (1.4 ± 0.1 mm (+)). Chitinase activity was again not observed in any tested temperatures (Figure 2, Table 1). Temperature significantly affected the extracellular enzymatic activities of *V. parahaemolyticus* (p-value < 0.05, Supplementary Table 2 B).

Investigating the effects of oxygen on extracellular enzymatic activities of *V. parahaemolyticus*

Under the limited oxygen presence, most of the tested enzyme activities were low or even not observed. Caseinase activity declined considerably from $1.10 \pm$

Table 1: Effect of pH, temperature and oxygen level on the extracellular enzymatic activity of *Vibrio parahaemolyticus* XN9. Tested pHs included 7.5, 8, 8.5 and 9. Activity was expressed via EA value in millimeter (mm). Tested temperatures included 25, 30 and 35°C. Oxygen conditions included shaking overnight culture followed by plate testing without AnaeroPack® (Nor-Nor), shaking overnight culture followed by plate testing with AnaeroPack (Nor-Li), static overnight culture with Oxygen absorber package followed by plate testing without AnaeroPack (Li-Nor), and static overnight culture with Oxygen absorber package followed by plate testing without AnaeroPack (Li-Li). Activity was expressed via EA value in millimeter (mm).

	Standard:	pH	Temperature			Oxygen condition		
			25oC	35oC	Nor-Nor	Nor-Li	Li-Nor	Li-Li
Caseinase	0.82 ± 0.17 (+)	7.5 3.55 ± 0.25 (++)	0.67 ± 0.10 (+)	0.85 ± 0.13 (+)	1.10 ± 0.31 (+)	0.46 ± 0.06 (+)	0.33 ± 0.13 (+)	-
Lecithinase	2.96 ± 0.13 (++)	8.0 2.85 ± 0.14 (++)	1.10 ± 0.15 (+)	1.87 ± 0.15 (+)	2.56 ± 0.19 (++)	2.92 ± 0.08 (++)	1.94 ± 0.18 (+)	1.62 ± 0.29 (+)
Gelatinase	0.89 ± 0.15 (+)	9.0 2.72 ± 0.12 (++)	0.86 ± 0.14 (+)	1.37 ± 0.25 (+)	1.59 ± 0.11 (+)	1.50 ± 0.21 (+)	1.27 ± 0.20 (+)	1.18 ± 0.13 (+)
Lipase	1.4 ± 0.1 (+)	8.0 1.77 ± 0.25 (+)	2.67 ± 0.22 (++)	2.25 ± 0.23 (++)	2.33 ± 0.23 (++)	2.20 ± 0.22 (++)	1.69 ± 0.14 (+)	1.17 ± 0.20 (+)
Chitinase	-	7.5 -	-	-	-	-	-	-

Table 2: Two-way ANOVA in analyzing the effect of A) four pH levels (7.5, 8.0, 8.5 and 9.0); B) three temperatures (25, 30 and 35°C); C) four tested oxygen conditions (Nor-Nor; Nor-Li; Li-Nor and Nor-Li) on the enzymatic activities of *V. parahaemolyticus*

A)							
Source of Variation	SS	df	MS	F	p-value	F crit	
Sample	50.3623567	4	12.5905892	335.719203	3.4326E-30	2.60597495	
Columns	2.31145833	3	0.77048611	20.5444701	3.2315E-08	2.8387454	
Interaction	23.97035	12	1.99752917	53.26271	1.0043E-20	2.0034594	
Within	1.50013333	40	0.03750333				
Total	78.1442983	59					
B)							
Source of Variation	SS	df	MS	F	p-value	F crit	
Sample	27.6018133	4	6.90045333	324.982104	3.1206E-24	2.68962757	
Columns	0.35015111	2	0.17507556	8.24531659	0.00140067	3.3158295	
Interaction	7.94242667	8	0.99280333	46.7568289	7.5498E-15	2.26616327	
Within	0.637	30	0.02123333				
Total	36.5313911	44					
C)							
Source of Variation	SS	df	MS	F	p-value	F crit	
Sample	42.5853233	4	10.6463308	253.032224	8.0542E-28	2.60597495	
Columns	5.012045	3	1.67068167	39.7072292	4.5515E-12	2.8387454	
Interaction	2.85693	12	0.2380775	5.65840761	1.4569E-05	2.0034594	
Within	1.683	40	0.042075				
Total	52.1372983	59					

0.31 mm (+) in the case of both fully oxygenated culture and testing to 0 mm (-) in case of both limited oxygen culture and testing. Similar trend was seen in case of lipase and gelatinase, with enzyme activity decreased markedly about 49% from 2.33 ± 0.23 mm (++) to 1.17 ± 0.20 mm (+) and 25% from 1.59 ± 0.11 (+) to 1.18 ± 0.13 mm (+) respectively. In case of lecithinase, this enzyme activity expressed in most tested conditions with lowest activity (1.62 ± 0.29 mm (+)) in case of both limited oxygen culture and testing and highest activity (2.92 ± 0.08 mm (++) in case of oxygenated overnight culture followed by limited oxygen testing condition. No activity of chitinase was

observed in any case (Figure 3, Table 1). Under four tested oxygen conditions, the activity of tested extracellular enzymes was different significantly (p -value < 0.05, Table 2 C).

DISCUSSION

Our data indicated that the production of extracellular enzymes in *V. parahaemolyticus* was highly affected by environmental factors. In inappropriate conditions, the production of some enzymes can be minimized to undetected levels such as gelatinase in case of pH 7.5 or caseinase in case of limited oxygen condition. On the other hand, some enzymes, such as

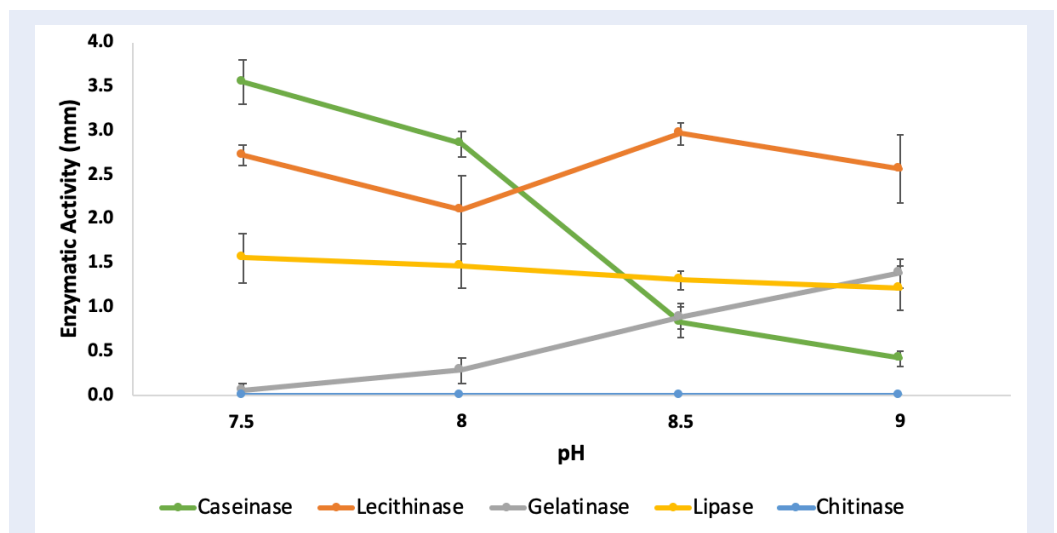


Figure 1: Extracellular enzymatic activity of *Vibrio parahaemolyticus* under different culturing pHs. No activity of chitinase was observed in any tested culturing pH.

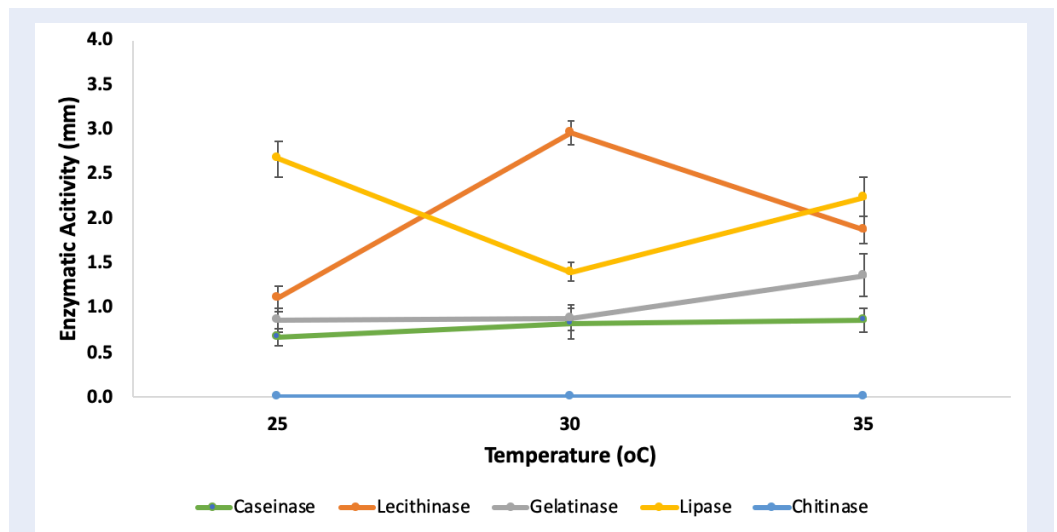


Figure 2: Extracellular enzymatic activity of *Vibrio parahaemolyticus* under different culturing temperatures. No activity of chitinase was observed in any tested culturing temperature.

lecithinase seemed to be constantly and strongly produced in most conditions tested in our study. The strong production of lecithinase was observed not only in *V. parahaemolyticus* but also in other *Vibrio* species^{17,19-21}. We did not detect chitinase activity in any tested conditions. Chitinase is a typical virulence factor of marine bacteria that can breakdown glycosidic bonds in the chitin of shrimp and other marine organisms²². The absence of chitinase activity indicated that this AHPND strain may not utilize chitinase attack aquatic crustaceans or chitinase might not

be induced *in vitro*.

Regarding pH, our data showed that while most of the tested enzymes showed the highest activity at pH 7.0, gelatinase only expressed its activity at alkaline conditions. This is in agreement with previous studies showing that *V. parahaemolyticus* had a high rate of hydrolysis of gelatin in alkaline environments^{23,24}. Gelatinase, together with lecithinase and protease, are constantly expressed in most disease-causing *Vibrio* species, particularly *V. parahaemolyticus* strains^{21,25}.

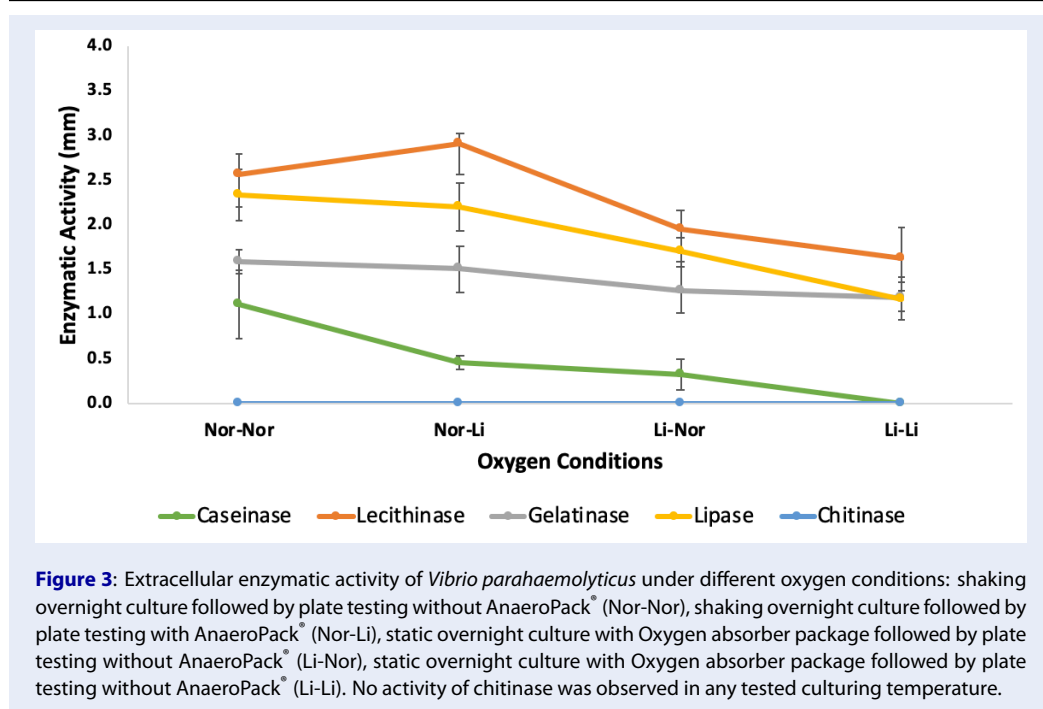


Figure 3: Extracellular enzymatic activity of *Vibrio parahaemolyticus* under different oxygen conditions: shaking overnight culture followed by plate testing without AnaeroPack® (Nor-Nor), shaking overnight culture followed by plate testing with AnaeroPack® (Nor-Li), static overnight culture with Oxygen absorber package followed by plate testing without AnaeroPack® (Li-Nor), static overnight culture with Oxygen absorber package followed by plate testing without AnaeroPack® (Li-Li). No activity of chitinase was observed in any tested culturing temperature.

Temperature is a well-known factor that affects the growth of *V. parahaemolyticus*. It was shown that the minimal growth temperature of *V. parahaemolyticus* was 13°C, and its optimal growth temperature was 30°C^{14,26}. However, for extracellular enzyme production, the optimal temperature was varied for different types of enzymes. Gelatinase, for examples was found to express the highest activity in *Vibrio* species at 24°C²³. In our study, the optimal temperature was 35°C for caseinase, gelatinase and lipase, and 30°C for lecithinase.

V. parahaemolyticus, like other *Vibrio* species, are facultative anaerobe. Its growth is only hindered by strict anaerobic conditions but not limited oxygen condition. Some *in vivo* environmental study even showed that the number of *V. parahaemolyticus* in low oxygen marine water was higher than in high oxygen samples²⁷. Our data showed a decrease in the activity of extracellular enzymes for all tested enzymes, of which caseinase was the most affected one. No activity of caseinase was found when *V. parahaemolyticus* was cultured and tested in limited oxygen conditions. It was in agreement with a previous study showing that the production of proteolytic enzymes was negatively affected by a low dissolved oxygen level²⁸.

CONCLUSION

pH, temperature, and oxygen condition are essential factors affecting not only the growth of *V. para-*

haemolyticus as previously shown (14) but also its extracellular enzyme activity. No culturing condition resulted in the highest activity for all extracellular enzymes was found. However, warm temperature (30/35°C), mildly alkaline pH (pH 8.0), and fully oxygenated condition could increase the overall extracellular enzymatic activity of *V. parahaemolyticus*, thus increase its potential virulence.

LIST OF ABBREVIATIONS

- V. parahaemolyticus*: *Vibrio parahaemolyticus*
- AHPND: Acute Hepatopancreatic Necrosis Disease
- BHI: Brain Heart Infusion
- EA: Enzyme activity
- EMS: Early Mortality Syndrome
- TCBS: Thiosulfate-citrate-bile salts-sucrose agar

COMPETING INTERESTS

The author(s) declare that they have no competing interests

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REFERENCES

1. Broberg CA, Calder TJ, Orth K. *Vibrio parahaemolyticus* cell biology and pathogenicity determinants. *Microbes Infect*;2011(13(12-13)):992-1001. PMID: 21782964. Available from: <https://doi.org/10.1016/j.micinf.2011.06.013>.

2. Hedberg CW. Foodborne illness acquired in the United States. *Emerg Infect Dis* Author reply 9-40. 2011;17(7):1338. PMID: 21762617. Available from: <https://doi.org/10.3201/eid1707.110019>.
3. Wu G, Yuan Q, Wang L, et al. Epidemiology of foodborne disease outbreaks from 2011 to 2016 in Shandong Province, China. *Medicine* (Baltimore). 2018;97(45):e13142. PMID: 30407341. Available from: <https://doi.org/10.1097/MD.000000000013142>.
4. Park K, Mok JS, Kwon JY, Ryu AR, Kim SH, Lee HJ. Food-borne outbreaks, distributions, virulence, and antibiotic resistance profiles of *Vibrio parahaemolyticus* in Korea from 2003 to 2016: a review. *Fisheries and Aquatic Sciences*. 2018;21(1):3. Available from: <https://doi.org/10.1186/s41240-018-0081-4>.
5. Sirikharin R, Taengchaiyaphum S, Sanguanrut P, Chi TD, et al. Characterization and PCR Detection Of Binary, Pir-Like Toxins from *Vibrio parahaemolyticus* Isolates that Cause Acute Hepatopancreatic Necrosis Disease (AHPND) in shrimp. *PLoS One*. 2015;10(5):e0126987. PMID: 26017673. Available from: <https://doi.org/10.1371/journal.pone.0126987>.
6. Lai HC, Ng TH, et al. Pathogenesis of acute hepatopancreatic necrosis disease (AHPND) in shrimp. *Fish Shellfish Immunol*. 2015;47(2):1006–1014. PMID: 26549178. Available from: <https://doi.org/10.1016/j.fsi.2015.11.008>.
7. Nunan L, Lightner D, Pantoja C, Gomez-Jimenez S. Detection of acute hepatopancreatic necrosis disease (AHPND) in Mexico. *Dis Aquat Organ*. 2014;111(1):81–86. PMID: <https://doi.org/10.3354/dao02776>. Available from: 25144120.
8. Yang YT, Chen IT, Lee CT, Chen CY, et al. Draft Genome Sequences of Four Strains of *Vibrio parahaemolyticus*, Three of Which Cause Early Mortality Syndrome/Acute Hepatopancreatic Necrosis Disease in Shrimp in China and Thailand. *Genome Announc*. 2014;2(5). PMID: 25189578. Available from: <https://doi.org/10.1128/genomeA.00816-14>.
9. Pena LD, Cabillon NA, Catedral DD, Amar EC, Usero RC, Monotilla WD, et al. Acute hepatopancreatic necrosis disease (AHPND) outbreaks in *Penaeus vannamei* and *P. monodon* cultured in the Philippines. *Dis Aquat Organ*. 2015;116(3):251–254. PMID: 26503780. Available from: <https://doi.org/10.3354/dao02919>.
10. Lee CT, Chen IT, Yang YT, Ko TP, et al. The opportunistic marine pathogen *Vibrio parahaemolyticus* becomes virulent by acquiring a plasmid that expresses a deadly toxin. *Proc Natl Acad Sci U S A*. 2015;112(34):10798–10803. PMID: 26261348. Available from: <https://doi.org/10.1073/pnas.1503129112>.
11. Kodama H, Moustafa M, Ishiguro S, Mikami T, Izawa H. Extracellular virulence factors of fish *Vibrio*: relationships between toxic material, hemolysin, and proteolytic enzyme. *Am J Vet Res*. 1984;45(10):2203–2207.
12. Miyoshi S. Extracellular proteolytic enzymes produced by human pathogenic *Vibrio* species. *Front Microbiol*. 2013;4:339. PMID: 24302921. Available from: <https://doi.org/10.3389/fmicb.2013.00339>.
13. Glenn AR. Production of extracellular proteins by bacteria. *Annual Review of Microbiology*. 1976;30(1):41–62. PMID: 791074. Available from: <https://doi.org/10.1146/annurev.mi.30.100176.000353>.
14. Anh PTL, Khang LQ, Thuc NT, Chau DNP, Nguyen TTH. Optimizing conditions for *Vibrio parahaemolyticus* culture and preservation. 7th International Conference on the Development of Biomedical Engineering in Vietnam ; Ho Chi Minh City. Springer. 2018;.
15. Nagpure A, Choudhary B, Gupta RK. Chitinases: in agriculture and human healthcare. *Crit Rev Biotechnol*. 2014;34(3):215–232. PMID: 23859124. Available from: <https://doi.org/10.3109/07388551.2013.790874>.
16. Vermelho AB, Meirelles MN, Lopes A, Petinate SD, Chaia AA, Branquinha MH. Detection of extracellular proteases from microorganisms on agar plates. *Mem Inst Oswaldo Cruz*. 1996;91(6):755–760. PMID: 9283660. Available from: <https://doi.org/10.1590/S0074-02761996000600020>.
17. Fiore AE, Michalski JM, Russell RG, Sears CL, Kaper JB. Cloning, characterization, and chromosomal mapping of a phospholipase (lecithinase) produced by *Vibrio cholerae*. *Infect Immun*. 1997;65(8):3112–3117. PMID: 9234762. Available from: <https://doi.org/10.1128/IAI.65.8.3112-3117.1997>.
18. Ohishi K, Murase K, Ohta T, Etoh H. Cloning and sequencing of a chitinase gene from *Vibrio alginolyticus* H-8. *J Biosci Bioeng*. 2000;89(5):501–505. Available from: [https://doi.org/10.1016/S1389-1723\(00\)89106-9](https://doi.org/10.1016/S1389-1723(00)89106-9).
19. Beleneva IA, Maslennikova EF, Magarlamov TY. Physiological and Biochemical Characteristics of the Halophilic Bacteria *Vibrio parahaemolyticus* and *V. alginolyticus* Isolated from Marine Invertebrates of Peter the Great Bay, Sea of Japan. *Russian Journal of Marine Biology*. 2004;30(2):96–100. Available from: <https://doi.org/10.1023/B:RUMB.0000025985.38429.11>.
20. Costa RA, Conde-Amorim LM, Araujo RL, Fernandes-Vieira RH. Multiple enzymatic profiles of *Vibrio parahaemolyticus* strains isolated from oysters. *Rev Argent Microbiol*. 2013;45(4):267–270. Available from: [https://doi.org/10.1016/S0325-7541\(13\)70035-X](https://doi.org/10.1016/S0325-7541(13)70035-X).
21. Bunpa S, Sermwittayawong N, Uddhakul V. Extracellular Enzymes Produced by *Vibrio alginolyticus* Isolated from Environments and Diseased Aquatic Animals. *Procedia Chemistry*. 2016;18:12–17. Available from: <https://doi.org/10.1016/j.proche.2016.01.002>.
22. Oyeleye A, Normi YM. Chitinase: diversity, limitations, and trends in engineering for suitable applications. *Biosci Rep*. 2018;38(4). PMID: 30042170. Available from: <https://doi.org/10.1042/BSR20180323>.
23. Weimer MS, Morita RY. Temperature and hydrostatic pressure effects on gelatinase activity of a *Vibrio* sp. and partially purified gelatinase. *Z Allg Mikrobiol*. 1974;14(8):719–725. PMID: 4619521. Available from: <https://doi.org/10.1002/jobm.19740140810>.
24. Stack MS, Gray RD. The effect of pH, temperature, and D2O on the activity of porcine synovial collagenase and gelatinase. *Arch Biochem Biophys*. 1990;281(2):257–263. Available from: [https://doi.org/10.1016/0003-9861\(90\)90441-Z](https://doi.org/10.1016/0003-9861(90)90441-Z).
25. Beshiru A, Igbinosa EO. Characterization of extracellular virulence properties and biofilm-formation capacity of *Vibrio* species recovered from ready-to-eat (RTE) shrimps. *Microb Pathog*. 2018;119:93–102. PMID: 29654902. Available from: <https://doi.org/10.1016/j.micpath.2018.04.015>.
26. Kim YW, Lee SH, Hwang IG, Yoon KS. Effect of temperature on growth of *Vibrio parahaemolyticus* [corrected] and *Vibrio vulnificus* in flounder, salmon sashimi and oyster meat. *Int J Environ Res Public Health*. 2012;9(12):4662–4675. PMID: 23330227. Available from: <https://doi.org/10.3390/ijerph9124662>.
27. Davis BJK, Jacobs JM, Davis MF, Schwab KJ, DePaola A, Curriero FC. Environmental Determinants of *Vibrio parahaemolyticus* in the Chesapeake Bay. *Appl Environ Microbiol*. 2017;83(21). PMID: 28842541. Available from: <https://doi.org/10.1128/AEM.01147-17>.
28. Wiersma M, Harder W. A continuous culture study of the regulation of extracellular protease production in *Vibrio* SA1. *Antonie van Leeuwenhoek*. 1978;p. 44. PMID: 582093. Available from: <https://doi.org/10.1007/BF00643217>.