CHEMICAL CONSTITUENTS OF THE LICHEN PARMOTREMA TSAVOENSE (KROG & SWINSCOW) KROG & SWINSCOW (PARMELIACEAE)

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ABSTRACT

Two major depsidones protocetraric acid (1), 8'-O-methylprotocetraric acid (2), virensic acid (3), two aliphatic acids (+)-prasorediosic acid (4), (+)-vinaprasorediosic acid A (5), and along with common lichen metabolites atranorin (6), methyl haematommate (7), methyl β -orsellinate (8), methyl orsellinate (9), zeorin (10) were isolated from the lichen Parmotrema tsavoense (Krog & Swinscow) Krog & Swinscow. Their chemical structures were established by 1D NMR, 2D NMR, high resolution ESI–MS spectroscopic analysis and comparison with those reported in the literatures. The lichen Parmotrema tsavoense has not been studied on phytochemistry.

Keywords: Parmotrema tsavoense, depsidone, aliphatic acid, monocyclic compound.

TÓM TẮT

Thành phần hóa học của loài địa y Parmotrema tsavoense (Krog & Swinscow) Krog & Swinscow (Parmeliaceae)

Hai hợp chất depsidone chính protocetraric acid (1), 8'-O-methylprotocetraric acid (2), virensic acid (3), hai acid béo (+)-prasorediosic acid (4), (+)-vinaprasorediosic acid A (5), và cùng với các hợp chất địa y phổ biến khác atranorin (6), methyl haematommate (7), methyl β -orsellinate (8), methyl orsellinate (9), zeorin (10) đã được cô lập từ loài địa y Parmotrema tsavoense (Krog & Swinscow) Krog & Swincow. Cấu trúc hoá học của chúng được xác định bằng các phương pháp phổ nghiệm cũng như so sánh với các tài liệu tham khảo. Loài địa y Parmotrema tsavoense chưa được nghiên cứu về hóa thực vật.

Từ khóa: Parmotrema tsavoense, depsidone, aliphatic acid, monocyclic compound.

1. Introduction

Depsidones and γ -lactone aliphatic acids are bioactive lichen metabolites (Huneck S. 1997).⁶ They possess the antivirus and enzyme inhibitory activities according to Boustie & Grube (2007),¹ Boustie *et al.* (2010),² Muller (2001).¹⁴ The previous phytochemical studies of the lichens *Parmotrema* genera growing in Vietnam indicate that they contain various depsidones and γ -lactone aliphatic acids (Huynh B. L. C., 2014, 2016).^{8,9} *Parmotrema tsavoense* (Krog & Swinscow) Krog & Swinscow has not been studied on phytochemisty.

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Figure 1. Chemical structures of 1-10

In this paper, we reported the isolation of ten known compounds protocetraric acid (1), 8'-O-methylprotocetraric acid (2), virensic acid (3), (+)-prasorediosic acid (4), (+)-vinaprasorediosic acid A (5), atranorin (6), methyl haematommate (7), methyl β -orsellinate (8), methyl orsellinate (9), zeorin (10) from the lichen *Parmotrema tsavoense*. Their chemical structures were elucidated by spectroscopic data analysis and comparison with those reported in the literature.

2. Experimental

General experimental procedures

The NMR spectra were measured on a Bruker Avance III (500 MHz for ¹H NMR and 125 MHz for ¹³C NMR) and Varian Mercury-400 Plus NMR (400 MHz for ¹H NMR and 100 MHz for ¹³C NMR) spectrometers with TMS as internal standard. Proton chemical shifts were referenced to the solvent residual signal of CDCl₃ at $\delta_{\rm H}$ 7.26, of CD₃COCD₃ at $\delta_{\rm H}$ 2.05, of DMSO- d_6



Figure 2. Parmotrema tsavoense on rock

at $\delta_{\rm H}$ 2.50. The ¹³C–NMR spectra were referenced to the central peak of CDCl₃ at $\delta_{\rm C}$ 77.1, of CD₃COCD₃ at $\delta_{\rm C}$ 29.4, of DMSO- d_6 at $\delta_{\rm C}$ 39.5. The HR–ESI–MS were recorded on a Bruker micrOTOF Q-II. TLC was carried out on precoated silica gel 60 F₂₅₄ or silica gel 60 RP–18 F₂₅₄S (Merck) and spots were visualized by spraying with 30% H₂SO₄ solution followed by heating. Gravity column chromatography was performed with Silica gel 60 (0.040–0.063 mm, Himedia).

Plant material

Parmotrema tsavoense (Krog & Swinscow) Krog & Swinscow was collected on the surface of rocks on Ta Cu mountain, Binh Thuan province (August-September 2012). Its scientific name was determined by Dr. Wetchasart Polyiam, Lichen Research Unit, Department of Biology, Faculty of Science, Ramkhamhaeng University, Bangkok, Thailand. A voucher specimen (No US-B027) was deposited in the herbarium of the Department of Organic Chemistry, University of Science.

Extraction and isolation

The clean, air-dried and ground material (1350 g) was extracted by methanol at ambient temperature, and the filtrated solution was concentrated under reduced pressure. While the methanolic solution was being evaporated, a precipitate (79.7 g) occurred and was filtered off. The rest of the solution was evaporated to dryness to obtain a crude methanol extract (249.8 g). This crude extract was applied to normal phase silica gel column chromatography, eluted with the solvent system of hexaneethyl acetate (9:1) to afford fraction P1 (9.9 g). Consecutive elution of the column with the same solvent system but increasing polarity (8:2, 7:3, 6:4, 5:5, 4:6) yielded five fractions, P2 (2.8 g), P3 (3.3 g), P4 (3.1 g), P5 (16.1 g), and P6 (9.9 g), respectively. Finally, the remaining residue was eluted with ethyl acetate-methanol in the ratios (9:1) and (0:10), respectively, to afford two fractions, P7 (5.1 g) and M (80.1 g). The precipitate was washed many times with acetone to afford compound 1 (19.8 g) and a washed solution (48.1 g). A part of the washed solution (1.1 g) was evaporated to dryness and applied to column chromatography to afford compound 2 (29.9 mg). A part of the extract **P1** (1.0 g) was applied to silica gel column chromatography, eluted with hexane-ethyl acetate-acetic acid (9:1:0.02) to give four compounds, 6 (295.7 mg), 7 (98.9 mg), 8 (199.1 mg), and 9 (99.8 mg). Fraction P3 (3.3 g) was rechromatographed, eluted with hexane-ethyl acetate-acetic acid (9:1:0.02) to give three fractions, P3.1 (0.1 g), P3.2 (0.8 g), and P3.3 (1.1 g). Fraction P3.1 was rechromatographed, eluted with hexane-ethyl acetate-acetic acid (9:1:0.02) to afford two compounds 3 (3.7 mg) and 10 (19.7 mg). Fraction P3.3 was purified by column chromatography, eluted with hexane-chloroform-ethyl acetate (1:1:1) to afford compound 4 (10.9 mg). Fraction P5 (16.1 g) was subjected to silica gel column chromatography with hexane-ethyl acetate-acetic acid (9:1:0.02) as eluent to obtain four fractions, P5.1 (1.1 g) and P5.2–5.4 (10.7 g). Applying fraction P5.1 to silica gel column chromatography yielded two fractions, **P5.1.1** (0.2 g) and **P5.1.2** (0.2 g). Fraction **P5.1.2** was further chromatographed to afford compound **5** (4.8 mg).

• **Protocetraric acid (1):** White amorphous powder. The ¹H- (500 MHz) and ¹³C-NMR (125 MHz) data (DMSO- d_6): see Table 1. HMBC correlations: see Figure 3. These spectroscopic data were suitable with those reported in the literature.³

• 8'-O-Methylprotocetraric acid (2): White amorphous powder. The ¹H- (500 MHz) and ¹³C-NMR (125 MHz) data (DMSO- d_6): see Table 1. HMBC correlations: see

Figure 3. These spectroscopic data were suitable with with those reported in the literatures.⁶

• Virensic acid (3): White amorphous powder. The ¹H- (400 MHz) and ¹³C-NMR (100 MHz) data (DMSO- d_6): see Table 1.. These spectroscopic data were suitable with those reported in the literature.⁶

• (+)-**Prasorediosic acid** (4): White amorphous powder. $[\alpha]_{D}^{23}$ + 439 (*c* 0.13, ethanol). HR-ESI-MS, negative mode: m/z 381.2265 [M-H]⁻ (calcd. for C₂₁H₃₄O₆-H, 381.2277). The ¹H- (500 MHz) and ¹³C-NMR (125 MHz) data (Acetone- d_6): see Table 2. These spectroscopic data were suitable with with those reported in the literature.⁹

• (+)-Vinaprasorediosic acid A (5): White amorphous powder. + 59 (*c* 0.28, ethanol). HR-ESI-MS, positive mode: m/z 417.2610 [M+Na]⁺ (calcd for C₂₃H₃₈O₅-H, 417.2617). The ¹H- (500 MHz) and ¹³C-NMR data (125 MHz) (DMSO-*d*₆): see Table 2. These spectroscopic data were suitable with with those reported in the literature.⁹

• Atranorin (6): Colorless needles (acetone). The ¹H-NMR (500 MHz)data (CDCl₃) were suitable with with those reported in the literature. ^{4,13} Methyl haematommate (7): Colorless needles (chloroform). The ¹H-NMR (500 MHz) data (CDCl₃) were suitable with with those reported in the literature. ^{4,12} Methyl β -orsellinate (8): Pale-green crystals (chloroform). The ¹H-NMR (500 MHz) data (Acetone-*d*₆) were suitable with with those reported in the literature. ^{4,7} Methyl orsellinate (9): White needles (chloroform). The ¹H-NMR (500 MHz) data (Acetone-*d*₆) were suitable with with those reported in the literature. ^{4,7} Methyl orsellinate (9): White needles (chloroform). The ¹H-NMR (500 MHz) data (Acetone-*d*₆) were suitable with those reported in the literature. ^{4,11} Zeorin (10): White amorphous powder. The ¹H-NMR (400 MHz) data (CDCl₃) were suitable with with those reported in the literature. ^{5,10} See Supporting information.

3. **Results and discussion**

Compound **1** was isolated as an amorphous powder. Its ¹H and ¹³C spectra revealed the presence of one aromatic methine ($\delta_{\rm H}$ 6.83, $\delta_{\rm C}$ 117.0), one aldehyde group ($\delta_{\rm H}$ 10.59, $\delta_{\rm C}$ 191.7), two methyls ($\delta_{\rm H}$ 2.43, $\delta_{\rm C}$ 21.3, C-9; $\delta_{\rm H}$ 2.40, $\delta_{\rm C}$ 14.3, C-9'), one hydroxymethylene group ($\delta_{\rm H}$ 4.60, $\delta_{\rm C}$ 52.9), two carboxyl groups ($\delta_{\rm C}$ 170.1, 163.9), and eleven aromatic quaternary carbons. From these data, **1** was presumed to be a depsidone. In the A-ring, the HMBC spectrum showed cross peaks of protons H-5 and 6-CH₃ to carbons C-6 and C-1. Moreover, protons H-5 and H-8 also correlated to C-3 and C-4. Thus, the structure of the A-ring was determined. In the B-ring, the hydroxymethylene showed the ³J correlation to carbon C-2' and the second methyl group also showed the ⁴J correlation to this carbon in the HMBC spectrum. Accordingly, the B-ring was elucidated (Fig. 3). Comparison of NMR data of **1** with the ones in the literature (Brandao L. F. G. et al., 2013)³ confirmed that **1** was protocetraric acid.

Compound 2 was isolated as a white amorphous powder. Comparison of the 1Dand 2D-NMR data of 2 and 1 (as shown in Table 1 and Figure 3) indicated that they were analogous, except for the presence of an additional methoxy group at $\delta_{\rm H}$ 3.22. Besides, this methoxy group showed a cross peak to C-8' on the basis of HMBC correlations. This indicated that the hydroxymethylene group was methyl etherificated. Comparison of the NMR data of 2 with those in the literature (Huneck S., 1997)⁶ indicated that 2 was 8'-O-methylprotocetraric acid.

	1 ^a		2^{a}		3 ^b	
Ν	$\delta_{ m H}$	δ_{C}	$\delta_{ m H}$	$\delta_{ m C}$	$\delta_{ m H}$	δ_0
1		112.4		112.5		112.3
2		161.2		161.5		161.3
3		111.8		112.5		111.8
4		163.8		164.4		163.8
5	6.83 (s)	117.0	6.84 (s)	117.6	6.83 (s)	117.0
6		152.0		152.2		152.0
7		163.9		164.6		164.0
8	10.59 (s)	191.7	10.58 (s)	192.0	10.59 (s)	191.7
9	2.43(s)	21.3	2.42(s)	21.8	2.43(s)	21.4
1'		116.6		116.5		115.7
2'		155.0		155.9		155.3
3'		118.6		116.3		115.1
4'		144.5		146.0		144.7
5'		141.7		142.6		141.8
6'		129.4		131.4		127.6
7'		170.1		170.8		170.8
8'	4.60(s)	52.9	4.50(s)	62.8	2.14(s)	9.3
9'	2.40(s)	14.3	2.40(s)	15.0	2.40(s)	14.3
8'-OCH ₃		115.6	3.22(s)	57.9		

Table 1. NMR data (DMSO-d₆) of 1-3



Figure 3. HMBC correlations of 1 and 2

^a recorded in 500 MHz for ¹H NMR, 125 MHz for ¹³C NMR ^b recorded in 400 MHz for ¹H NMR, 100 MHz for ¹³C NMR

Compound **3** was isolated as a white amorphous solid. The NMR spectral data of **3** contained all signals as found in **1**, except for the replacement of the hydroxymethylene group H₂-8' ($\delta_{\rm H}$ 4.60, $\delta_{\rm C}$ 52.9) as in **1** by the methyl group H₃-8' ($\delta_{\rm H}$ 2.14, $\delta_{\rm C}$ 9.3). This resulted in the slightly differences of ¹³C chemical shift values in the B-ring. Comparison of NMR data of **3** with the ones in the literature (Huneck S., 1996)⁶ confirmed that **3** was virensic acid.

Compound 4 was isolated as a white amorphous powder. The ¹H NMR spectrum revealed one *multiplet* oxymethine ($\delta_{\rm H}$ 5.15), one *doublet* methyl at $\delta_{\rm H}$ 2.03. Moreover, the ¹H NMR spectrum exhibited one broad *singlet* signal at $\delta_{\rm H}$ 1.28 with high intensity together with one *triplet* methylene at $\delta_{\rm H}$ 2.18 characteristic of the γ -lactone skeleton with a long chain alkyl group.^{6,9} The chemical shift of the *triplet* methylene at $\delta_{\rm H}$ 2.18 in accordance with the presence of the ¹³C carboxyl signal at $\delta_{\rm C}$ 174.3 indicated the presence of the fragment -CH₂–CH₂–COOH. These findings were consistent with the molecular formula, C₂₁H₃₄O₆. Accordingly, **4** was elucidated to be prasorediosic acid (Huynh B.L.C, 2016, Huneck S., 1997).^{6,9} Moreover, the specific rotation of **4** is dextrorotary, similarly to that of (+)-prasorediosic acid (Huynh B.L.C, 2016),

indicating the (4R) configuration of 4. Consequently, 4 was determined to be (+)-prasorediosic acid.

Compound **5** was isolated as a white amorphous powder. Comparison of the NMR data between **5** and **4** indicated that they were very similar, except for the presence of the acetyl group at ($\delta_{\rm H}$ 2.04, $\delta_{\rm C}$ 28.9, 208.3, H₃C-C=O) and the absence of the *triplet* highfield methyl of **5** at $\delta_{\rm H}$ 0.87. This difference was supported by the molecular formula of **5**, C₂₃H₃₈O₅. Moreover, the NMR data as well as the specific rotation of **5** were very similar to (+)-vinaprasorediosic acid A (Huynh B.L.C, 2016),⁹ indicating that the absolute configuration of C-4 was (*R*). Accordingly, **5** was elucidated as (+)-vinaprasorediosic acid A.

Table 2. NMR data of 4 and 5									
.	4 ^a		5 ^b						
Ν	$\delta_{ m H}$ J (Hz)	$\delta_{ m C}$	Ν	$\delta_{ m H\!$	$\delta_{ m C}$				
1		172.4	1		172.7				
2		134.2	2		nd				
3		149.8	3		nd				
4	5.15 (<i>m</i>)	80.9	4	5.13 (m)	80.3				
5	2.03 (d, 2.0)	10.3	5	2.02 (<i>d</i> , 2.0)	10.2				
6		163.5	6		163.3				
7	1.55 (<i>m</i>), 1.20 (<i>m</i>)	31.6	7	1.42 (<i>m</i>)	31.3				
8	1.21 (<i>m</i>)	24.5	8	1.21 (<i>m</i>)	24.8				
9-17	1.21 (<i>m</i>)	28.0-29.0	9-19	1.21 (<i>m</i>)	28.029.0				
18	1.48 (<i>m</i>)	23.1	20	1.52 (<i>m</i>)	23.1				
19	1.99 (<i>m</i>)	24.1	21	2.37 (t, 7.0)	42.6				
20	2.18 (<i>t</i> , 7.0)	33.4	22		208.3				
21		174.3	23	2.04 (s)	28.9				

^a recorded in acetone- d_6 , ^b recorded in DMSO- d_6 , nd: not determined

4. Conclusion

Ten known compounds were isolated from the lichen *Parmotrema tsavoense* collected in Binh Thuan province. Two depsidones protocetraric acid (1), 8'-O-methylprotocetraric acid (2) were isolated as major components of the lichen. This is the first time these ten compounds are reported in *Parmotrema tsavoense*. Further studies on this lichen are in progress.

REFERENCES

- 1. Boustie J., Grube M. (2007), "Lichens a promising source of bioactive secondary metabolites", *Plant Genetic Resources*, **3**(2), 273-287.
- 2. Boustie J., Tomasi S. and Grube M. (2010), "Bioactive lichen metabolites: alpine habitats as an untapped source", *Phytochemistry Reviews*, **10**(3), 287-307.

- 3. Brandão L. F. G., Alcantara G. B., Matos M. de F. C., Bogo D., Freitas D. dos S., Oyama N. M., Honda N. K. (2013), "Cytotoxic evaluation of phenolic compounds from lichens against melanoma cells", *Chemical and Pharmaceutical Bulletin*, 61(2), 176–183.
- 4. Duong T. H., Huynh B. L. C., Ha X. P., Ton T. Q., Nguyen K. P. P. (2011), "Some phenolic compounds of the lichen Parmotrema planatilobatum (Hale) Hale (Parmeliaceae)", *Journal of Science and Technology Development*, 14(6), 5–10.
- 5. Duong T. H., Tran T. T. (2015), "Some hopanes and ergostanes from the lichen Parmotrema sancti-angelii (Lynge) Hale (Parmeliaceae)", *Journal of Science-University of Pedagogy*, 2(67), 13–20.
- 6. Huneck S., Yoshimura I. (1997), *Identification of lichen substances*, Springer Verlag Berlin.
- Hylands, P. J., Ingolfsdottir, K. (1985), "The isolation of methyl β-orsellinate from Stereocaulon alpinum and comments on the isolation of 4,6-dihydroxy-2-methoxy-3methylacetophenone from Stereocaulon species", *Phytochemistry*, 24(1), 127–129.
- 8. Huynh Bui Linh Chi (2014), "Chemical constituents of lichen Parmotrema prasorediosum collected in Vienam", A thesis for Doctor of Philosophy, *University of Science*, National University-Ho Chi Minh City.
- Huynh B. L. C., Duong T. H., Do T. M. L., Pinnock G. T., Pratt L. M., Yamamoto S., Watarai H., Tanahashi T., Nguyen K. P. P. (2016), "New γ-lactone carboxylic acids from the Lichen Parmotrema prasorediosum (Nyl.) Hale, Parmeliacea", *Records of Natural Products*, 10(3), 332-340.
- 10. Konig G. M., Wright A. D. (1999), "1H and 13C-NMR and biological activity investigations of four lichen-derived compounds", *Phytochemical Analysis*, 10, 279 284.
- 11. Lopes T. I. B., Coelho R. G., Yoshida N. C., Honda N. K. (2008), "Radicalscavenging activity of orsellinates", *Chemical and Pharmaceutical Bulletin*, 56, 1551-1554.
- 12. Marante F. J. T., Castellano A. G., Rosas F. E., Aguiar J. Q., Barrera J. B. (2003), "Identification and quantitation of allelochemicals from the lichen Lethariella canariensis: phytotoxicity and antioxidative activity", *Journal of Chemical Ecology*, 29, 2049–2071.
- Micheletti A. C., Beatriz A., Lima D. P. de, Honda N. K. (2009), "Chemical constituents of Parmotrema lichexanthonicum Eliasaro & Adler Isolation, structure modification and evaluation of antibiotic and cytotoxic activities", *Química Nova*, 32, 12–20.
- 14. Muller .K. (2001), "Pharmaceutically relevant metabolites from lichens", *Applied Microbiology and Biotechnology*, 56, 9-16.

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